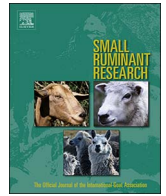


# TUTDoR

## Attainment of puberty in South African unimproved indigenous bucks.

Item Type	Article
Authors	Ramukhithi, Fhulufhelo Vincent;Nephawe, Khathutshelo Agree;Chokoe, Tlou Caswell;Matabane, Matshidiso Bailakaye;Mphaphathi, Masindi Lotus;Lehloenya, Khoboso Christina;Nedambale, Tshimangadzo Lucky
DOI	<a href="http://dx.doi.org/10.1016/j.smallrumres.2017.05.009">http://dx.doi.org/10.1016/j.smallrumres.2017.05.009</a>
Publisher	Elsevier
Rights	Attribution-NonCommercial-ShareAlike 4.0 International
Download date	2025-03-27 00:23:09
Item License	<a href="http://creativecommons.org/licenses/by-nc-sa/4.0/">http://creativecommons.org/licenses/by-nc-sa/4.0/</a>
Link to Item	<a href="https://hdl.handle.net/20.500.14519/1217">https://hdl.handle.net/20.500.14519/1217</a>



## Research Paper

## Attainment of puberty in South African unimproved indigenous bucks



Fhulufhelo Vincent Ramukhithi<sup>a,b</sup>, Khathutshelo Agree Nephawe<sup>b</sup>, Tlou Caswell Chokoe<sup>c</sup>,  
Matshidiso Bailakaye Matabane<sup>a</sup>, Masindi Lotus Mphaphathi<sup>a</sup>, Khoboso Christina Lehloenyana<sup>d</sup>,  
Tshimangadzo Lucky Nedambale<sup>b,\*</sup>

<sup>a</sup> Agricultural Research Council, Germplasm Conservation and Reproductive Biotechnologies, Private Bag X 2, Irene, 0062, South Africa

<sup>b</sup> Tshwane University of Technology, Department of Animal Sciences, Private Bag X 680, Pretoria, 0001, South Africa

<sup>c</sup> Department of Agriculture, Forestry and Fisheries, Directorate: Farm Animal Genetic Resource, Private Bag X 250, Pretoria, 0001, South Africa

<sup>d</sup> University of Pretoria, Department of Animal and Wildlife Sciences, Private Bag X 20, Hatfield, 0028, South Africa

## ARTICLE INFO

## Keywords:

Puberty attainment  
Indigenous bucks  
Sexual behaviour activities  
Testosterone

## ABSTRACT

The purpose of the study was to examine attainment of puberty in South African unimproved indigenous bucks (unimproved indigenous bucks). A total of 24 unimproved indigenous bucks aged between 118–123 days (4 months) with a mean body weight of  $8.1 \pm 0.3$  kg were observed for puberty attainment. Their sexual behaviour activities, body weight and scrotal circumference were recorded every two weeks until the onset of puberty. Blood samples were collected every month to evaluate testosterone concentration. Semen samples were collected when the bucks started to show sexual behaviour activities. For continuous and categorical variables, least-square mean and percentage were used to summarise the data, respectively. Bucks started to show sexual behaviour activities at the age of 5.5 months. At the age of 5.5 months, none of the bucks ejaculated semen with the sperm cells. However, at the age of 6 months, 22 bucks ejaculated semen and the semen characteristics observed were good and above the acceptable standards. In conclusion, unimproved indigenous bucks attained puberty at the age of 6 months.

## 1. Introduction

Goat meat consumption is increasing worldwide due to its high nutritional value, cholesterol composition and its leanness (Supakorn, 2009; Agga et al., 2011; Astro Awani, 2015; Tardiff, 2015). Production of goat meat needs to be increased to keep up with the growing demand (Sodiq, 2004; Sebei, 2005). This may be achieved through good reproductive performance. Moreover, in order to establish the appropriate selection criteria for young breeding goats, it is crucial to understand the reproductive functioning from the onset of puberty (Nishimura et al., 2000; Bezerra et al., 2009). Evaluation of bucks' puberty is a complex process that cannot be defined by single trait. Therefore, evaluation of age, body weight, scrotal circumference and gonadotropins levels are used to determine puberty attainment (Bezerra et al., 2009). Their combined effects cannot be separated when semen-producing capacity is evaluated (Bester, 2006).

Puberty in bucks is attained when they start to produce viable sperm cells that are capable of producing pregnancy (Daramola et al., 2007; Bezerra et al., 2009). During puberty attainment in bucks, the levels of follicle stimulating hormone (FSH) or spermatogenic stimulating hormone (SSH) and luteinizing hormone (LH) or interstitial cell stimulat-

ing hormone (ICSH) in the testes increase (Bester, 2006). Luteinizing hormone stimulates the leydig cells to produce testosterone hormone (Farshad et al., 2012), which then initiates the onset of puberty and sperm cells development (Bezerra et al., 2009). Then, FSH stimulates spermiogenesis in the presence of testosterone, which is responsible for sexual behaviour activities (nosing, mounting, nudging, bleating, pawing, flehmen, licking, pelvic thrusts and penile erection) which are correlated to sexual maturity (Nishimura et al., 2000; Farshad et al., 2012) and maintenance of ideal conditions for spermiogenesis and semen ejaculation (Bester, 2006).

Puberty attainment varies and it is primarily influenced by body weight, breed type, birth season and management systems (Daramola et al., 2007; Delgadillo et al., 2007; Farshad et al., 2012). In bucks, puberty has been documented to be reached from the age of 5 months (Souza et al., 2011). Bucks that reach puberty earlier have higher reproductive capacity and shorter generation gap (Bezerra et al., 2009; Bitto and Egbunike, 2012). Irrespective of goat being seasonal breeders, different breeds differ on the rate in which they attain puberty (Nishimura et al., 2000; Bester, 2006; Aguirre et al., 2007). Therefore, gathering of information including puberty attainment in different goat breeds, is crucial for good reproductive management of a herd, such as

\* Corresponding author.

E-mail address: [nedambaletl@tut.ac.za](mailto:nedambaletl@tut.ac.za) (T.L. Nedambale).

selecting of males capable of making females pregnant (Nishimura et al., 2000; Bezerra et al., 2009). However, in unimproved indigenous goats and other indigenous goats in developing countries, there is little information on their reproductive performance including puberty attainment (Webb et al., 1998; Bitto and Egbunike, 2012). Moreover, much attention in unimproved indigenous goats has been paid to the adult goats than in young goats (Matshaba, 2010; Ramukhithi et al., 2011, 2015; Bopape et al., 2015). The purpose of the study was therefore to examine attainment of puberty in unimproved indigenous bucks.

## 2. Materials and methods

The study was approved by ethic committees of the Tshwane University of Technology (REC2012/10/019-2) and Agricultural Research Council (APIEC15/044). A total of 24 South African unimproved indigenous bucks also known as unimproved indigenous bucks aged around 4 months (range from 118 to 123 days) with a mean body weight of  $8.1 \pm 0.3$  kg were used. Unimproved indigenous goats are mostly small sized animals, tolerate harsh environmental conditions, parasites and diseases, and are able to survive on poor quality grazing when compared to exotic breeds (Ramsay and Donkin, 2000). The bucks were weaned at 4 months of age. The first evaluation was done after weaning (4 months of age) and the last evaluation was done at the age of 6 months. Moreover, the bucks were grazing together on natural pasture (*Kikuyu – Pennisetum clandestinum*) and drinking water was provided *ad libitum* through metal drinking trough.

To determine puberty attainment of the experimental bucks, sexual behaviour activities, body weight (Nishimura et al., 2000) and testosterone concentration (Santiago-Moreno et al., 2005) were evaluated. For sexual behaviour activities evaluation (visual observation), the bucks were given access for 20 min to teaser does which were on heat every second week until they reached puberty (6 bucks versus 1 doe per session). The sexual behaviour activities observed were; nosing, nudging, pawing, bleating, flehmen, licking, mounting, penile erection and pelvic thrusts (Nishimura et al., 2000). For collection of the sexual behaviour activities data, each buck was observed and counted based on number of the times it showed any sexual behavioural activity. Body weight and scrotal circumference were also determined every second week using a weighing scale and flexible tape, respectively.

For determination of testosterone concentration, blood samples were collected once a month until bucks reached puberty. The blood samples were then centrifuged at  $1500 \times g$  for 10 min ( $25^\circ\text{C}$ ). Blood serum were harvested and stored in 1 mL tubes at  $-20^\circ\text{C}$  until analysed. A liquid chromatography tandem mass spectrometry (LC-MS/MS) method was modified and validated for the detection of seven steroids including testosterone (Guan et al., 2010). In this method, testosterone was extracted from serum with methyl *tert*-butyl ether (MTBE) and analysed using multiple reaction monitoring (MRM) acquisition mass spectrometric detection in positive electron mode using electrospray ionization (ESI). The linear range of the method was from 1 to 50 (parts per billion) ppb. The samples were then quantified using matrix-matched standard addition calibration curves. The limit of quantification for the method was 1 ppb.

Semen samples were collected following the onset of sexual behaviour activities (5.5 months) using an electro-ejaculator (Ramsem, South Africa). Prior to semen collection, hair around the sheath was shaved and the prepuce was cleaned with a sterile paper towel containing 70% ethanol for the prevention of contamination. Before insertion of the rectal probe, it was washed with 70% ethanol and lubricated with a lubrication jelly. The probe was inserted and placed in the rectum above the accessory sex glands in order to stimulate them (Dombo, 2002). Semen samples were collected in pre-warmed ( $37^\circ\text{C}$ ) 15 mL graduated tubes. Four levels of 30 voltage were applied when the bucks were lying on their side (Ramukhithi, 2011). From 24 unimproved indigenous bucks that showed sexual behaviour

activities, only 22 bucks responded to electro-ejaculation stimulation and ejaculated semen.

Following semen collection, semen samples were then evaluated for volume by reading the measurements on the collection tubes (Yamashiro et al., 2006) and pH using a pH meter (HANNA Instruments®, South Africa). Sperm cell motility was determined using a Sperm Class Analyser® (Microptic S.L, Barcelona), whereby 500  $\mu\text{L}$  of Tris and 10  $\mu\text{L}$  of semen were mixed in a 1 mL graduated tube and incubated for 5 min at  $37^\circ\text{C}$ . After incubation, 10  $\mu\text{L}$  of extended semen was placed on a pre-warmed microscopic slide ( $37^\circ\text{C}$ ), mounted with a cover slip and examined ( $\times 10$ ) under a phase contrast microscope (Ramukhithi, 2011). Sperm cell concentration was determined with a spectrophotometer (Jenway, United Kingdom). A square cuvette was filled with 3 mL of sodium citrate solution and placed in a spectrophotometer for at least 30 s. Raw semen (15  $\mu\text{L}$ ) was added in a square cuvette containing the sodium citrate solution, again placed in a spectrophotometer in order to read the absorbance. The absorbance read was used to determine the final sperm cell concentration with the aid of a formula ( $201 \times 25.97 \times \text{absorbance} - 0.3$ ). The final sperm cell concentration was recorded in millions per millilitre (Seshoka, 2015).

For evaluation of acrosome integrity, sperm cell viability, morphology and abnormalities, samples stained with nigrosin-eosin stain were evaluated under a fluorescent microscope (Olympus, Japan) and 200 sperm cells per slide were counted. Live sperm cells and the sperm cells that have non-reacted acrosomes did not absorb stain (fluorescence), while dead sperm cells and the sperm cells that have reacted acrosomes absorbed stain and become purple in colour (Samper, 2000). Live sperm cells were further evaluated for morphology and abnormalities. Abnormalities were categorised as primary, secondary and tertiary abnormalities (Loskutoff and Crichton, 2001). For evaluation of membrane integrity, a hypo-osmotic swelling (HOS) test was used and the samples were evaluated under a phase contrast microscope ( $400\times$ ) and 200 sperm cells per slide were counted. Sperm cells with swollen and coiled tail were considered intact (Naing et al., 2010).

The data were analysed using Statistical Analysis Software (SAS) (1999), Version 9.2. Sexual behaviour activities data for all the bucks were summarised as percentages. Analysis of variance (ANOVA) and least significant difference (LSD) were applied in order to analyse the influence of bucks' age on the body weight and scrotal circumference. Body weight, scrotal circumference, testosterone concentration ( $\alpha = 0.05$ ) and semen characteristics data were summarised as least square mean  $\pm$  standard error. The Pearson correlation was done to show the relationship between body weight, scrotal circumference, testosterone concentration and semen characteristics of unimproved indigenous bucks.

## 3. Results

Sexual behaviour activities of unimproved indigenous bucks are illustrated in Table 1. No sexual behaviour activities were observed at the age of 4–5 months. However, at the age of 5.5 months, unimproved indigenous bucks started to show sexual behaviour activities. The highest proportion of sexual behaviour activity at the age of 5.5 months was mounting (50%) and the lowest proportion was pelvic thrusts (4%). At the age of 6 months, the highest proportion of sexual behaviour activity observed was licking (83%) and the lowest proportion was penile erection (17%). The sexual behaviour activities had increased by 50% from the age of 5.5–6 months.

Body weight, scrotal circumference and testosterone concentration of unimproved indigenous bucks are illustrated in Table 2. The body weights of unimproved indigenous bucks ranges from  $8.1 \pm 0.3$  to  $14.6 \pm 0.5$  kg at the age of 4–6 months. Moreover, body weights of unimproved indigenous bucks observed from the age of 4–6 months were significant different ( $p < 0.05$ ). Scrotal circumferences at ages of 5 ( $15.5 \pm 0.4$  cm), 5.5 ( $16.7 \pm 0.4$  cm) and 6 months

**Table 1**  
Sexual behaviour activities of unimproved indigenous bucks (%) (n = 24).

Sexual behaviour activities	Age				
	4 months	4.5 months	5 months	5.5 months	6 months
Nosing	0	0	0	17	46
Nudging	0	0	0	21	25
Mounting	0	0	0	50	50
Bleating	0	0	0	29	46
Pawing	0	0	0	21	25
Flehmen	0	0	0	21	75
Licking	0	0	0	8	83
Pelvic thrusts	0	0	0	4	25
Penile erection	0	0	0	25	17
<b>Average</b>	0	0	0	22	44

**Table 2**  
Body weight, scrotal circumference and testosterone concentration of unimproved indigenous bucks (LS-mean ± se) (n = 24).

Age (months)	Body weight (kg)	Scrotal circumference (cm)	Testosterone (ng/mL)
4	8.1 ± 0.3 <sup>a</sup>	12.9 ± 0.4 <sup>a</sup>	< 1.0
4.5	9.6 ± 0.4 <sup>b</sup>	14.1 ± 0.4 <sup>a</sup>	
5	11.4 ± 0.4 <sup>c</sup>	15.5 ± 0.4 <sup>b</sup>	2.1 ± 0.03 <sup>a</sup>
5.5	12.8 ± 0.4 <sup>d</sup>	16.7 ± 0.4 <sup>c</sup>	
6	14.6 ± 0.5 <sup>e</sup>	18.1 ± 0.4 <sup>d</sup>	4.5 ± 0.2 <sup>b</sup>

<sup>a,b</sup>Values with different superscripts within the same column differ significantly (p < 0.05).

(18.1 ± 0.4 cm) were significantly different (p < 0.05). Testosterone concentration at the age of 4 months was below 1 ng/mL. Testosterone concentration was higher at the age of 6 months (4.5 ± 0.2 ng/mL) when compared to 5 months (2.1 ± 0.03 ng/mL) (p < 0.05).

Semen characteristics of unimproved indigenous bucks are illustrated in Table 3. At the age of 5.5 months, none of the unimproved

**Table 3**  
Semen characteristics of unimproved indigenous bucks collected at the age of 6 months (n = 22).

Characteristics		LS-mean ± se		
Semen	Volume (mL)		0.6 ± 0.04	
		pH	7.0 ± 0.1	
	Concentration (X 10 <sup>9</sup> /mL)		2.9 ± 0.3	
Sperm cell	Motility	Progression (%)	TM	78.7 ± 3.7
			PM	42.8 ± 4.9
			NPM	35.9 ± 3.2
			Static	21.3 ± 3.7
			Rapid	28.9 ± 5.3
	Velocity (%)	Average values of velocity parameters	Medium	30.5 ± 3.7
			Slow	33.6 ± 9.4
			VCL (µm/s)	85.5 ± 7.2
			VSL (µm/s)	62.5 ± 7.6
			VAP (µm/s)	69.4 ± 7.5
	Vitality (%)	Acrosome Membrane	LIN (%)	63.8 ± 3.5
			STR (%)	73.9 ± 3.5
			WOB (%)	80.3 ± 2.9
			Intact	96.7 ± 0.6
			Intact	82.3 ± 1.8
Viability (%)	Live		84.9 ± 2.8	
		Morphology (%)	92.1 ± 1.3	
Abnormalities (%)	Primary		1.6 ± 0.2	
		Secondary	3.5 ± 0.6	
		Tertiary	2.8 ± 0.5	

TM = total motility, PM = progressive motility, NPM = non-progressive motility, VCL = curvilinear velocity, VSL = straight-line velocity, VAP = average path velocity, LIN = linearity, STR = straightness and WOB = wobble.

indigenous bucks did ejaculate semen with the sperm cells. However, at the age of 6 months, 22 unimproved indigenous bucks ejaculated, and the semen characteristics observed were good and above the acceptable standards.

The Pearson correlation between body weight, scrotal circumference, testosterone concentration and semen characteristics of unimproved indigenous bucks are illustrated in Table 4. Body weight did not show any correlation with TM and static sperm cells. On the other hand, body weight had a positive correlation with intact acrosome and membrane (p < 0.01). However, body weight had a negative correlation with slow velocity (p < 0.01). Scrotal circumference had a positive correlation with intact acrosome (p < 0.01). Testosterone concentration did not show any correlation with sperm cell viability and primary abnormalities.

#### 4. Discussion

When the current study was compared with the previous studies: unimproved indigenous bucks had a lower body weight at the age of 4 (8.1 kg) and 6 months (14.6 kg) when compared to other indigenous bucks in the previous studies at the same age. For example, Tokara bucks were weighing 10.6 and 15.8 kg at the age of 4 and 6 months, respectively (Nishimura et al., 2000). Moreover, Boer goat bucks were weighing 16.3 kg at the age of 3 months (Bezerra et al., 2009). The scrotal circumference that was observed at the age of 4 months remained the same at the age of 4.5 months, and it started to increase from 5 to 6 months of age. The results of the current study are in line with the literature, as it has been documented that body weight and scrotal circumference increase with age (Delgadillo et al., 2007; Bezerra et al., 2009). The scrotal circumference of 18.1 cm observed in unimproved indigenous bucks which were 6 months old were lower when compared to Boer goat scrotal circumference of 21 cm which had the same age (Bezerra et al., 2009). The differences between the current study and previous studies might be due to climate, latitude, hormonal concentrations (Bezerra et al., 2009; Farshad et al., 2012), breed and nutritional differences (Gebre, 2007; Akpa et al., 2013).

In the current study, bucks started to show sexual behaviour activities at the age of 5.5 months, with mounting being the highest, followed by licking at the age of 6 months. This is in agreement with the literature as it had showed that mounting of does by bucks is a common and first activity that can be used to evaluate puberty of bucks when introduced to the does (Nishimura et al., 2000; Suyadi, 2012). Generally, sexual behaviour activities had increased exhibition with age of the bucks. In contrast, Suyadi (2012) reported that age of bucks has no influence on the sexual behaviour activities.

At the age of 4 months, unimproved indigenous bucks' testosterone concentration was below 1 ng/mL. This confirmed that at that age those bucks were still immature. Testosterone concentration in the current study increased with the age of the bucks. This also agrees with the previous studies (Bezerra et al., 2009; Souza et al., 2011). Moreover, testosterone concentration of 4.5 ng/mL in unimproved indigenous bucks was high and comparable to Markhoz matured bucks (2–3 years) testosterone concentration (4.8 ng/mL) (Farshad et al., 2012). On the other hand, testosterone concentration at the age of 6 months was even higher when compared to the previous study in Boer goat bucks at the age of 6 months (1.9 ng/mL) (Bezerra et al., 2009). The increase of testosterone concentration with age in the current study is also in agreement with the literature (Farshad et al., 2012). Moreover, testosterone levels in the current study is in agreement with Souza et al. (2011) who indicated that testosterone concentration ranged from 0.4 to 5.4 ng/mL at the age of 5–9.5 months. The reactivation of leydig cells was documented to be associated with the multiplication of germ cells, which is a crucial physiological event on achieving puberty in bucks. The full development of the seminiferous tubules coincides with increase in testosterone concentration (Nishimura et al., 2000).

At the age of 5.5 months, none of the unimproved indigenous bucks

**Table 4**

The Pearson correlation between body weight, scrotal circumference, testosterone concentration and semen characteristics of unimproved indigenous bucks at the age of 6 months.

Characteristics		Body weight (kg)	Scrotal circumference (cm)	Testosterone (ng/mL)	
Semen	Volume (mL)	0.320	0.152	-0.344	
	pH	-0.054	0.070	0.054	
Sperm cell	Concentration (X 10 <sup>9</sup> /mL)	0.013	-0.194	-0.181	
	Progression (%)	TM	0	-0.162	0.362
		PM	0.049	-0.114	0.037
		NPM	-0.081	-0.013	0.371
		Static	0	0.161	-0.362
		Rapid	-0.027	-0.296	-0.008
	Velocity (%)	Medium	-0.106	0.090	0.222
		Slow	-0.658**	-0.389	0.157
		Average values of velocity parameters	VCL (µm/s)	0.004	-0.065
		VSL (µm/s)	-0.357	-0.215	0.080
		VAP (µm/s)	0.133	0.074	0.029
		LIN (%)	-0.119	0.122	0.039
		STR (%)	-0.091	0.119	-0.098
		WOB (%)	0.011	0.189	0.052
	Vitality (%)	A/intact	0.543**	0.485**	-0.196
		M/intact	0.439**	0.124	-0.028
	Viability (%)	Live	0.089	-0.122	0
		Morphology (%)	Normal	-0.137	-0.008
	Abnormalities (%)	Primary	-0.080	0.207	0
Secondary		0.111	-0.094	0.029	
Tertiary		0.125	-0.214	0.278	

\*\*Significant at  $p < 0.01$ , TM = total motility, PM = progressive motility, NPM = non-progressive motility, VCL = curvilinear velocity, VSL = straight-line velocity, VAP = average path velocity, LIN = linearity, STR = straightness, WOB = wobble, A = acrosome and M = membrane.

did ejaculate semen with the sperm cells. However, at the age of 6 months 22 bucks ejaculated, and the semen characteristics observed were within acceptable limits (Ramukhithi et al., 2011). This shows that the puberty has been reached (Souza et al., 2011). Moreover, this also shows that the semen characteristics of unimproved indigenous bucks were significantly influenced by age like in other goat breeds. Semen production with acceptable characteristics at the age of 6 months was reported to be due to a significant development of seminiferous tubules and sertoli cells (Souza et al., 2011). However, the onset of semen production in the current study (6 months) was late when compared to Tokara bucks (4.5 months) (Nishimura et al., 2000) and West African Dwarf bucks (5 months) (Bitto and Egbunike, 2012). Semen characteristics in the current study were better when compared to Boer goat bucks, which were even older (Suyadi, 2012).

Body weight showed positive relationship with intact acrosome and membrane. This might be an indication that the animals that are well fed and their body condition is improved, as a result some of their semen characteristics are also improved (Akpa et al., 2013). On the other hand, scrotal circumference showed positive relationship with intact acrosome. For the other semen characteristics that were positive or negative with body weight, scrotal circumference and testosterone, their relationships were low and insignificant. However, this contradicts with the literature, whereby most of semen characteristics were reported to have positive and significant relationship with body weight and scrotal circumference (Webb et al., 2004; Pepper-Yowell, 2011).

Unimproved indigenous bucks attained puberty at the age of 6 months with the average body weight of 14.6 kg and scrotal circumference of 18.1 cm. However, these were lower when compared to body weight of 25.7 kg and scrotal circumference of 21.1 cm reported in Anglo-Nubian bucks that reached puberty at the age of 5 months. This show that breed, environment, management systems and genetic type influences attainment of puberty (Daramola et al., 2007; Souza et al., 2011).

## 5. Conclusion

Unimproved indigenous bucks attained puberty at the age of 6

months with a significant increase in body weight, scrotal circumference and testosterone concentration.

## Conflict of interest

There is no conflict of interest to declare.

## Acknowledgements

The authors would like to thank Agricultural Research Council, Tshwane University of Technology, Department of Agriculture, Forestry and Fisheries, and Council for Scientific and Industrial Research – Southern African Science Service Centre for Climate Change and Adaptive Land Management for their contributions.

## References

- Agga, G.E., Udala, U., Regassa, F., Wudie, A., 2011. Body measurements of bucks of three goat breeds in Ethiopia and their correlation to breed, age and testicular measurements. *Small Rumin. Res.* 95, 133–138.
- Aguirre, V., Orihuela, A., Vazquez, R., 2007. Effect of semen collection frequency on seasonal variation in sexual behaviour, testosterone, testicular size and semen characteristics of tropical hair rams (*Ovis aries*). *Trop. Anim. Health Prod.* 39, 271–277.
- Akpa, G.N., Ambali, A.L., Suleiman, I.O., 2013. Body conformation, testicular and semen characteristics as influenced by age, hair type and body condition of Red Sokoto goat. *NY Sci J.* 6 (7), 44–58.
- Astro Awani, 2015. Goat Meat Good for Your Heart. <http://english.astroawani.com/lifestyle/goat-meat-good-your-heart-15581.com> (Accessed 8th June 2015).
- Bester, N., 2006. Effect of Different Dietary Energy Levels on Productive and Reproductive Traits in Dorper Rams. MSc Dissertation. University of Free State, Bloemfontein.
- Bezerra, F.Q.G., Aguiar Filho, C.R., Freitas Neto, L.M., Santos Junior, E.R., Chaves, R.M., Azevedo, E.M.P., Santos, M.H.B., Lima, P.F., Oliveira, M.A.L., 2009. Body weight, scrotal circumference and testosterone concentration in young Boer goat males born during the dry or rainy seasons. *S. Afr. J. Anim. Sci.* 39 (4), 30–306.
- Bitto, I.I., Egbunike, G.N., 2012. The semen characteristics of pubertal West African dwarf bucks. *Pertanika J. Trop. Agric. Sci.* 35 (2), 191–197.
- Bopape, M.A., Lehloeny, K.C., Chokoe, T.C., Nedambale, T.L., 2015. Comparison of electro ejaculator and artificial vagina on semen collection from South African indigenous goat following assessment by Computer aided sperm analysis. *Open. J. Anim. Sci.* 5, 210–218.
- Daramola, J.O., Adeloye, A.A., Fatoba, T.A., Soladoye, A.O., 2007. Induction of puberty in

- West African Dwarf buck-kids with exogenous melatonin. *Livestock Res. Rural Dev.* 19 (127).
- Delgadillo, J.A., De Santiago-Miramontes, M.A., Carrillo, E., 2007. Season of birth modifies puberty in female and male goats raised under subtropical conditions. *Animal* 1 (6), 858–864.
- Dombo, M.H., 2002. Seasonal Effect on Semen Quality of Gorno Altai and South African Indigenous Goats. MSc Dissertation. University of Pretoria, Pretoria.
- Farshad, A., Yousefi, A., Moghaddam, A., Khalili, B., 2012. Seasonal changes in serum Testosterone, LDH concentration and semen characteristics in Markhoz goats. *Asian Australas. J. Anim. Sci.* 25 (2), 189–193.
- Gebre, Y.M., 2007. Reproductive Traits in Ethiopian Male Goats, with Special Reference to Breed and Nutrition. Doctoral Thesis. Acta Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Guan, F., Uboh, C.E., Soma, L.R., You, Y., Liu, Y., Li, X., 2010. High-throughput UHPLC/MS/MS method for the detection, quantification and identification of fifty-five anabolic and androgenic steroids in equine plasma. *J. Mass. Spectrom* 45, 1270–1279.
- Loskutoff, N.M., Crichton, E.G., 2001. Standard Operating Procedures for Genome Resource Banking. The Bill and Berniece Brewcock Center for Conservation and Research. Omaha's Henry Doorly Zoopp. 1–16.
- Matshaba, B., 2010. Characterisation and Cryopreservation of South African Unimproved Indigenous Goat Semen. MSc Dissertation. University of the Free State, Bloemfontein.
- Naing, S.W., Wahid, H., Azam, K.M., Rosnis, Y., Zuki, A.B., Kazhal, S., Bukar, M.M., Thein, M., Kyaw, T., San, M.M., 2010. Effects of sugar on characteristics of Boer goat semen after cryopreservation. *Anim. Reprod. Sci.* 122, 23–28.
- Nishimura, S., Okano, K., Yasukouchi, K., Gotoh, T., Tabata, S., Iwamoto, H.K., 2000. Testis developments and puberty in the male Tokara (Japanese native) goat. *Anim. Reprod. Sci.* 64, 127–131.
- Pepper-Yowell, A.R., 2011. The Use of Computer Assisted Semen Analysis to Predict Fertility in Holstein Bulls. Msc. Dissertation. Colorado State University, Colorado.
- Ramsay, K.A., Donkin, E.F., 2000. A review of the current status of goat research and development in South Africa. In: *Proceedings of Regional Workshop on Goat Development in Southern Africa*. 31 July – 4 August 2000, Malawi, Mangochi. Available from: [https://www.google.co.in/search?q=www.+researchgate.net/.../237254983+A+review+of+the+current+s...pdf&ie=utf-8&oe=utf-8&gws\\_rd=cr&ei=WLYiWcndBsnuvATH3Y7QAw.com](https://www.google.co.in/search?q=www.+researchgate.net/.../237254983+A+review+of+the+current+s...pdf&ie=utf-8&oe=utf-8&gws_rd=cr&ei=WLYiWcndBsnuvATH3Y7QAw.com) (Accessed: 13th April 2014).
- Ramukhithi, F.V., Nedambale, T.L., Sutherland, B., Lehloeny, K.C., 2011. Cryopreservation of South African indigenous goat semen. *Afr. J. Biotechnol.* 10 (77), 17898–17902.
- Ramukhithi, F.V., Lehloeny, K.C., Kotze, A., Luther, I., Nephawe, K.A., Jonker, T., Chokoe, T.C., Maree, L., Nedambale, T.L., 2015. Comparison between South African unimproved indigenous and Tankwa goat semen characteristics. In: *Proceedings of the 48th South African Society of Animal Science Congress*. 21–23 September 2015, Empangeni, Kwazulu Natal Province, South Africa.
- Ramukhithi, F.V., 2011. Characterisation and Cryopreservation of South African Unimproved Indigenous and Boer Goat Semen. M.Tech Dissertation. Tshwane University of Technology, Pretoria.
- SAS Institute Inc, 1999. SAS/STAT User's Guide, Version 9, vol. 2 SAS Institute Inc, SAS Campus Drive, Cary, North Carolina 27513 (1st printing).
- Samper, C.J., 2000. Equine Breeding Management and Artificial Insemination, second ed. Saunders, Elsevier.
- Santiago-Moreno, J., Gomez-Brunet, A., Gonzalez-Bulnes, A., Toledano-Diaz, A., Malpau, B., Lopez-Sebastian, A., 2005. Differences in reproductive pattern between wild and domestic rams are not associated with inter-specific annual variations in plasma prolactin and melatonin concentrations. *Domest. Anim. Endocrinol.* 28, 416–429.
- Sebei, P.J., 2005. The Assessment of Some Factors Influencing the Survival of Kids in a Small-scale Communal Goat Production System. Msc. Dissertation. University of Pretoria, Pretoria.
- Seshoka, M.M., 2015. Quality of Frozen-thawed Nguni Bull Sperm Following Analysis Using the Computer Aided Sperm Analyser (CASA). M.Tech. Dissertation. Tshwane University of Technology, Pretoria.
- Sodiq, A., 2004. Doe Productivity of Kacang and Peranakan Etawah Goats and Factors Affecting Them in Indonesia. Doctoral Thesis. University of Kassel, Germany.
- Souza, C.E.A., Moura, A.A., Lima-Souza, A.C., Killian, G.J., 2011. Binding patterns of seminal plasma proteins on bovine epididymal and ejaculated sperm membrane. *J. Vet. Res. Anim. Sci.* 63 (3), 535–543.
- Supakorn, C., 2009. The important candidate genes in goats – a review. *Walailak J. Sci. Technol.* 6 (1), 17–36.
- Suyadi, S., 2012. Sexual behaviour and semen characteristics of young male Boer goats in tropical Condition: a case in Indonesia. *Int. J. Biol. Food. Vet. Agric. Eng.* 6 (6), 47–50.
- Tardiff, M., 2015. Goat Meat Nutrition Facts. <http://www.livestrong.com/article/330241-goat-meat-nutrition-facts/.com> (Accessed 18th June 2015).
- Research and training strategies for goat production systems in South Africa. In: Webb, E.C., Cronje, P.B., Donkin, E.F. (Eds.), *Proceedings of Goat Production in South Africa: Constraints and Opportunities Workshop*. November 22–26, Eastern Cape, South Africa. (Accessed 12th August 2012). <http://www.ais.up.ac.za/vet/goat/documents/hogsproceed.pdf>.
- Webb, E.C., Dombo, M.H., Roets, M., 2004. Seasonal variation in semen quality of Gorno Altai cashmere goats and South African indigenous goats. *S. Afr. J. Anim. Sci.* 34, 240–243.
- Yamashiro, H., Kumamoto, K., Wang, H., Yamashita, Y., Terada, T., 2006. Effect of semen collection in extender solution on the characteristics of goat spermatozoa. *J. Reprod. Dev.* 52 (3), 397–406.